

AMYRIS OF JAMAICA. NICOTINAMIDES OF AMYRIS PLUMIERI D.C., (RUTACEAE)

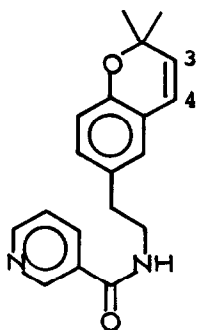
Basil A. Burke* and Helen Parkins

Department of Chemistry, University of the West Indies, Mona,
Kingston 7, Jamaica.

(Received in UK 9 May 1978; accepted for publication 25 May 1978)

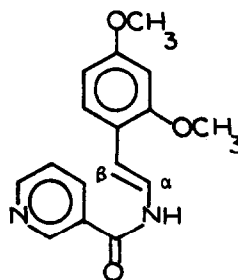
Unlike most rutaceous genera, Amyris¹ has attracted little attention of chemists. Recently however, there have been reports of coumarins in A. simplicifolia² and A. madrensis³, and upon re-examination the essential oil of A. balsamifera⁴ has yielded several sesquiterpenes. We now describe the formulation of two nicotinamides found among products from the benzene extract of the Jamaican variety⁵ of Amyris plumieri a plant which on botanical grounds was placed in the family Burseraceae⁶ but which has been reclassified as Rutaceae¹.

The chromene I, C₁₉H₂₀N₂O₂, m.p. 99-100°, was characterised on the basis of its spectral data. UV maxima at 224.5, 263.5 and 315 (log ε 4.43, 3.84, 3.32 respectively) nm indicated a chromene and other aromatic moiety, while IR bands at 3225 and 1653 cm⁻¹ suggested amide.



I.

II. 3,4,-dihydro



III.

IV. α,β -dihydro

The ^1H NMR (60 MHz) spectrum of I in CDCl_3 gave signals at δ 1.43 (6H, singlet) and 5.58 and 6.25 (each 1H, doublet, $J = 9$ Hz) for the hetero portion of the 2,2-dimethyl chromene. Signals at δ 2.81 (2H, triplet, $J = 7$ Hz), 3.67 (2H, quartet, $J = 7$ Hz, collapsing to triplet, 2 H, after shaking with D_2O) and 6.80 (1H, vanishing after exchange with D_2O) indicated the moiety $\text{ArCH}_2\text{CH}_2\text{NHCO-}$, while four low field protons, located at δ 7.52 - 8.83 were characteristic of a nicotinamide group⁷. The substitution pattern on the aryl portion of the chromene was revealed by signals at δ 6.65 (1H, doublet, $J = 8$ Hz), 6.78 (1H, doublet, $J = 2$ Hz) and 6.91 (1H, doublet of doublets, $J = 8, 2$ Hz).

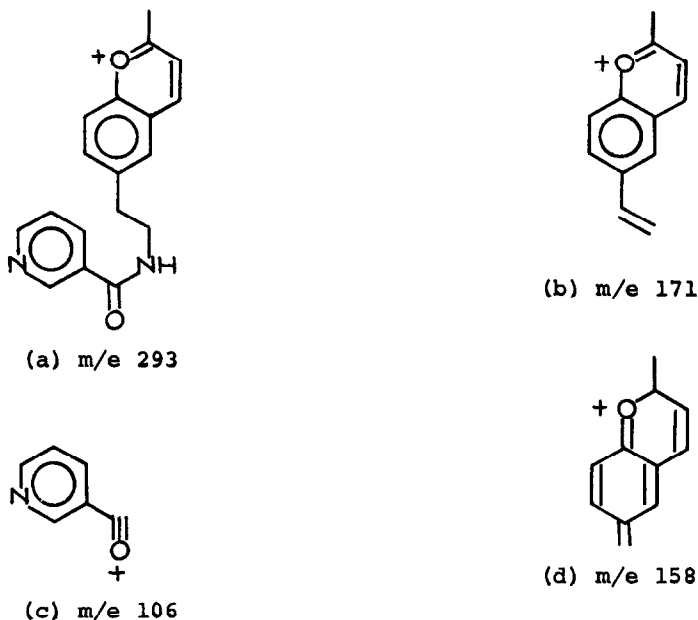


Figure I

Together with others, the fragments a - d (Figure I) obtained from mass spectral analysis of I supported these gross features.

Hydrogenation of I over 10% Pd/C gave the dihydroderivative II, $\text{C}_{19}\text{H}_{22}\text{N}_2\text{O}_2$, m.p. 125-128°; λ_{max} 226 and 264 (log ϵ 4.20 and 3.82 respectively) nm. The IR and ^1H NMR (60 MHz) spectra supported the structure II.

The structure I was finally confirmed to be a 6-substituted chromene by a biogenetic type synthesis via prenylation of the nicotinamide derivative of tyramine.

Compound III, $\text{C}_{16}\text{H}_{16}\text{N}_2\text{O}_3$, m.p. 159-160° had UV maxima at 265.5 and 366.6 (log ϵ 3.31 and 3.50 respectively) nm. IR bands at 3177, 1653 and 1605 cm^{-1} indicated amide and aryl moieties. Immediately obvious from the ^1H NMR (60 MHz) spectrum of III in CDCl_3 were the signals for the nicotinamide segment. In addition, there were signals at δ 6.47 (1H, doublet, $J = 14.5$ Hz) and 7.63 (1H,

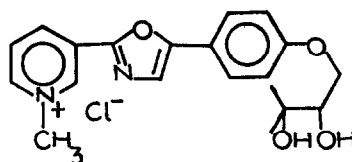
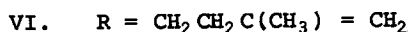
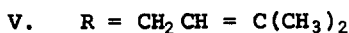
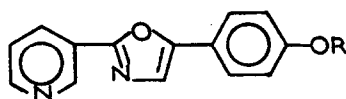
doublet of doublets, $J = 14.5, 10$ Hz, collapsing to a doublet $J = 14.5$ Hz upon shaking the sample with D_2O) for the vinyl protons of the group $ArCH = CH-NH-CO-$. One benzenoid proton partially overlapped with those of the heteroaromatic ring while the other two appeared as a multiplet at $\delta 6.54$.

Further confirmation of the styrylamide grouping was obtained upon hydrogenation of III with 10% Pd/C in ethanol to give IV, $C_{16}H_{18}N_2O_3$, m.p. 112-113°. The UV [215.5, 271 ($\log \epsilon$ 4.14 and 3.73 respectively) nm] and the IR (3344 and 1661 cm^{-1}) spectra of IV were consistent with the change, while the 1H NMR spectrum showed the signals for $ArCH_2CH_2NH$ CO- at $\delta 2.90$ (2H, triplet, $J = 6$ Hz), 3.72 (2H, quartet, $J = 6$ Hz, collapsing to a triplet, 2H, after shaking the sample with D_2O) and an exchangeable proton at $\delta 6.48$ (1H, broad multiplet).

Ozonolysis of III to yield 2,4-dimethoxybenzaldehyde and synthesis of IV from the latter compound confirmed the gross structure of III. The trans-geometry of the styryl linkage was based on the coupling constant ($J = 14.5$ Hz) of the olefinic protons.

The presence of the chromene and amide⁸ moieties in Amyris plumieri supports the botanical classification of the species as rutaceous.

This was confirmed by the discovery in the same plant of the masked nicotinamide *O*-dimethylallyl halfordinol, V, an oxazole, derivatives of which have been



VII.

found only in the Rutaceae. Compound V had m.p. 115-118°; the UV absorption [λ_{max} 250, 261 and 327.5 ($\log \epsilon$ 4.06, 4.03 and 4.40 respectively) nm] gave a bathochromic shift in acid to 261 and 346 ($\log \epsilon$ 4.21 and 4.24 respectively) nm, and IR bands were at 1616, 1605, 1582 and 822 cm^{-1} . The 1H NMR spectrum showed the presence of a 3-substituted pyridine (4 protons between $\delta 7.43$ and 9.47) a *p*-disubstituted benzenoid ring ($\delta 6.44, 8.44$, 2H each, doublets, $J = 9$ Hz) and a 3,3-dimethylallyl ether ($\delta 1.80$, 6H, broad singlet, 4.70, 2H doublet, $J = 6.5$ Hz coupled to a triplet, 1H, at $\delta 5.62$). The oxazole proton was a sharp singlet at $\delta 7.47$. These properties coincide with those reported by Dreyer⁹ for V, which was found in a mixture with *O*-isopentenyl isomer VI, and can be related to those of *N*-methyl halfordinium chloride VII¹⁰.

The compounds I and III are the first reported unmasked nicotinamides in Rutaceae.

REFERENCES AND NOTES

1. C.D. Adams, Flowering Plants of Jamaica, Robert MacLehose and Co. Ltd., The University Press, Glasgow, 1972, p. 388.
2. H.E. Cordova and L.E. Garelli, Phytochemistry, 13, 758, (1974).
3. X.A. Dominquez, G. Cano, I. Luna and A. Dieck, Phytochemistry, 16, 1090 (1977).
4. M. Rohmer. A.C. Schwartz and R. Anton, Phytochemistry, 16, 773 (1977).
5. A specimen of A. plumieri, Voucher # 34158, was deposited at the Herbarium of the Institute of Jamaica through the kindness of G. Proctor who collected the sample for us.
6. (a) R. Pernet, Lloydia, 35, 280 (1972); (b) H.L. Gerth van Wijk, A Dictionary of Plant Names, A. Asher and Co., Vaals-Amsterdam, 1971, p.77.
7. J.W. Emsley, J. Feeney and R.H. Sutcliffe, High Resolution Nuclear Magnetic Resonance Spectroscopy, Vol. 2, Pergamon, Oxford, 1965, p. 795.
8. (a) A. Chatterjee, M. Chakrabarty and A.B. Kundu, Aust. J. Chem., 28, 457 (1975); (b) idem. Chem. and Ind., 433 (1975); (c) F. Fish and P.G. Waterman, Taxon, 22, 177 (1973).
9. D.L. Dreyer, J. Org. Chem., 33, 3658 (1968).
10. W.D. Crow and J.H. Hodgkin, Aust. J. Chem., 17, 119 (1964).